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(54) Title: METHYLATION ALTERED DNA SEQUENCES AS MARKERS ASSOCIATED WITH HUMAN CANCER

(57) Abstract: There is disclosed (103) novel methylation-altered DNA sequences ("marker sequences") that have distinct methylation patterns in cancer, compared to normal tissue. In many instances, these marker sequences represent novel sequences not found in the GenBank data base, and none of these marker sequences have previously been characterized with respect to their methylation pattern in human cancers including, but not limited to those of bladder and prostate. These (103) sequences have utility as diagnosis, prognostic and therapeutic markers in the treatment of human cancer, and as reagents in kits for detecting methylated CpG-containing nucleic acids.

METHYLATION ALTERED DNA SEQUENCES AS MARKERS ASSOCIATED WITH HUMAN CANCER

5 Cross-Reference to Related Applications

This application claims priority to U.S. Patent Application Serial No. 09/699,243, filed October 27, 2000.

Technical Field of the Invention

The present invention relates to novel human DNA sequences that exhibit altered methylation patterns (hypermethylation or hypomethylation) in cancer patients. These novel methylation-altered DNA sequences are useful as diagnostic, prognostic and therapeutic markers for human cancer.

15 Background of the Invention

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The identification of early genetic changes in tumorigenesis is a primary focus in molecular cancer research. Characterization of the nature and pattern of cancer-associated genetic alterations will allow for early detection, diagnosis and treatment of cancer. Such genetic alterations in vertebrates fall generally into one of three categories: gain or loss of genetic material; mutation of genetic material; or methylation at cytosine residues in CpG dinucleotides within "CpG islands." Among these, DNA methylation is uniquely reversible, and changes in methylation state are known to affect gene expression (e.g., transcriptional initiation of genes where CpG islands located at or near the promoter region) or genomic stability.

Methylation of CpG dinucleotides within CpG islands. DNA, in higher order eukaryotic organisms, is methylated only at cytosine residues located 5' to guanosine residues in CpG dinucleotides. This covalent modification of the C-5 position of the cytosine base by the enzyme DNA (cytosine-5)-methyltransferase results in the formation of 5-methylcytosine (5-mCyt), and gives this base unique properties (e.g., susceptibility to undergo spontaneous deamination). This enzymatic conversion is the only epigenetic modification of DNA known to exist in vertebrates, and is essential for normal embryonic development (Bird, A.P., Cell 70:5-8, 1992; Laird & Jaenisch, Human Molecular Genetics 3:1487-1495, 1994; Li et al., Cell 69:915-926, 1992).

The presence of 5-mCyt at CpG dinucleotides has resulted in the 5-fold depletion of this sequence in the genome during the course of vertebrate evolution (Schroderet & Gartler, *Proc. Nat. Acad. Sci. USA* 89:957-961, 1992), presumably due to spontaneous deamination of 5-mCyt to Thymidine. Certain areas of the genome, however, do not show such depletion,

and are referred to as "CpG islands" (Bird, A.P., Nature 321:209-213, 1986; Gardiner-Garden & Frommer, J. Mol. Biol. 196:261-282, 1987). These CpG islands comprise only approximately 1% of the vertebrate genome, yet account for about 15% of the total number of genomic CpG dinucleotides (Antequera & Bird, Proc. Nat. Acad. Sci. USA 90:11995-11999, 1993). CpG islands contain the expected (i.e., the non-evolutionarily depleted) frequency of CpGs (with an Observed/Expected Ratio¹ >0.6), are GC-rich (with a GC Content² >0.5) and are typically between about 0.2 to about 1 kb in length.

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Methylation within CpG islands affects gene expression. CpG islands are located upstream of many housekeeping and tissue-specific genes, but may also extend into gene coding regions (Cross & Bird, Current Opinions in Genetics and Development 5:309-314, 1995; Larsen et al., Genomics 13:1095-1107, 1992). The methylation of cytosines within CpG islands in somatic tissues is believed to affect gene expression. Methylation has been inversely correlated with gene activity and may lead to decreased gene expression by a variety of mechanisms including inhibition of transcription initiation (Bird, A.P., Nature 321:209-213, 1986; Delgado et al., EMBO Journal 17:2426-2435, 1998), disruption of local chromatin structure (Counts & Goodman, Molecular Carcinogenesis 11:185-188, 1994; Antequera et al., Cell 62:503-514, 1990), and recruitment of proteins that interact specifically with methylated sequences and thereby directly or indirectly prevent transcription factor binding (Bird, A.P., Cell 70:5-8, 1992; Counts & Goodman, Molecular Carcinogenesis 11:185-188, 1994; Cedar, H., Cell 53:3-4, 1988). Many studies have demonstrated the effect of methylation of CpG islands on gene expression (e.g., the CDKN2A/p16 gene; Gonzalez-Zulueta et al., Cancer Research 55:4531-4535, 1995), but most CpG islands on autosomal genes remain unmethylated in the germline, and methylation of these islands is usually independent of gene expression. Tissue-specific genes are typically unmethylated in the respective target organs but are methylated in the germline and in non-expressing adult tissues, while CpG islands of constitutively expressed housekeeping genes are normally unmethylated in the germline and in somatic tissues.

Methylation within CpG islands affects the expression of genes involved in cancer. Data from a group of studies show the presence of altered methylation in cancer cells relative to non-cancerous cells. These studies show not only alteration of the overall genomic levels of DNA methylation, but also changes in the distribution of methyl groups. For example, abnormal methylation of CpG islands that are associated with tumor suppressor genes or oncogenes within a cell may cause altered gene expression. Such altered gene expression may provide a population of cells with a selective growth advantage and thereby result in selection of these cells to the detriment of the organism (i.e., cancer).

¹ Calculated as: [number of CpG sites / (number of C bases X number of G bases)] X band length for each fragment.

² Calculated as: (number of C bases + number of G bases) / band length for each fragment.

Insufficient correlative data. Unfortunately, the mere knowledge of the basic existence of altered methylation of CpG dinucleotides within CpG islands of cancer cells relative to normal cells, or of the fact that in particular instances such methylation changes result in altered gene expression (or chromatin structure or stability), is inadequate to allow for effective diagnostic, prognostic and therapeutic application of this knowledge. This is because only a limited number of CpG islands have been characterized, and thus there is insufficient knowledge, as to which particular CpG islands, among many, are actually involved in, or show significant correlation with cancer or the etiology thereof. Moreover, complex methylation patterns, involving a plurality of methylation-altered DNA sequences, including those that may have the sequence composition to qualify as CpG islands, may exist in particular cancers.

Therefore there is a need in the art to identify and characterize specific methylation altered DNA sequences, and to correlate them with cancer to allow for their diagnostic, prognostic and therapeutic application.

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Summary of the Invention

The present invention provides for a diagnostic or prognostic assay for cancer, comprising: obtaining a tissue sample from a test tissue; performing a methylation assay on DNA derived from the tissue sample, wherein the methylation assay determines the methylation state of a CpG dinucleotide within a DNA sequence of the DNA, and wherein the DNA sequence is a sequence selected from the group consisting of sequences of SEQ ID NOS:1-103, sequences having a nucleotide sequence at least 90% identical to sequences of SEQ ID NOS:1-103, CpG island sequences associated with sequences of SEQ ID NOS:1-103, CpG island sequences associated with sequences having a nucleotide sequence at least 90% identical to sequences of SEQ ID NOS:1-103, and combinations thereof, wherein the CpG island sequence associated with the sequence of the particular SEQ ID NO is that contiguous sequence of genomic DNA that encompasses at least one nucleotide of the particular SEQ ID NO sequence, and satisfies the criteria of having both a frequency of CpG dinucleotides corresponding to an Observed/Expected Ratio >0.6, and a GC Content >0.5; and determining a diagnosis or prognosis based, at least in part, upon the methylation state of the CpG dinucleotide within the DNA sequence. Preferably, the DNA sequence is a sequence selected from the group consisting of CpG island sequences associated with sequences of SEQ ID NOS:1-103, CpG island sequences associated with sequences having a nucleotide sequence at least 90% identical to sequences of SEQ ID NOS:1-103, and combinations thereof. Preferably, the DNA sequence is a sequence selected from the group consisting of CpG island sequences associated with sequences of SEQ ID NOS: 2, 4, 6, 7, 9-16, 19, 20, 22-33, 35-43, 48, 51-55, 59, 60, 64, 71, 76, 78-81, 84 and 87-90, and combinations thereof. Preferably, the methylation assay procedure is selected from the group

consisting of MethyLight, MS-SnuPE (methylation-sensitive single nucleotide primer extension), MSP (methylation-specific PCR), MCA (methylated CpG island amplification), COBRA (combined bisulfite restriction analysis), and combinations thereof. Preferably, the methylation state of the CpG dinucleotide within the DNA sequence is that of

hypermethylation, hypomethylation or normal methylation. Preferably, the cancer is selected from the group consisting of bladder cancer, prostate cancer, colon cancer, lung cancer, renal cancer, leukemia, breast cancer, uterine cancer, astrocytoma, glioblastoma, and neuroblastoma. Preferably, the cancer is bladder cancer, or prostate cancer.

The present invention further provides a kit useful for the detection of a methylated 10 CpG-containing nucleic acid comprising a carrier means containing one or more containers comprising: a container containing a probe or primer which hybridizes to any region of a sequence selected from the group consisting of SEQ ID NOS:1-103, and sequences having a nucleotide sequence at least 90% identical to sequences of SEQ ID NOS:1-103; and additional standard methylation assay reagents required to affect detection of methylated CpG-containing nucleic acid based on the probe or primer. Preferably, the additional 15 standard methylation assay reagents are standard reagents for performing a methylation assay from the group consisting of MethyLight, MS-SNuPE, MSP, MCA, COBRA, and combinations thereof. Preferably, the probe or primer comprises at least about 12 to 15 nucleotides of a sequence selected from the group consisting of SEQ ID NOS:1-103, and sequences having a nucleotide sequence at least 90% identical to sequences of SEQ ID 20 NOS:1-103.

The present invention further provides an isolated nucleic acid molecule comprising a methylated or unmethylated polynucleotide sequence selected from the group consisting of SEQ ID NO:1, SEQ ID NO:5, SEQ ID NO:6, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:13, SEQ ID NO:18, SEQ ID NO:24, SEQ ID NO:25, SEQ ID NO:32, SEQ ID NO:34, SEQ ID NO:37, SEQ ID NO:38, SEQ ID NO:42, SEQ ID NO:44, SEQ ID NO:51, SEQ ID NO:52, SEQ ID NO:62, SEQ ID NO:64, SEQ ID NO:65, SEQ ID NO:68, SEQ ID NO:69, SEQ ID NO:70, SEQ ID NO:71, SEQ ID NO:74, SEQ ID NO:76, SEQ ID NO:82, SEQ ID NO:83, SEQ ID NO:84, SEQ ID NO:86, SEQ ID NO:90, SEQ ID NO:92, SEQ ID NO:97, and SEQ ID NO:100. Preferably the nucleic acid is methylated. Preferably, the nucleic acid is unmethylated.

Detailed Description of the Invention

35 Definitions:

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"GC Content" refers, within a particular DNA sequence, to the [(number of C bases + number of G bases) / band length for each fragment].

"Observed/Expected Ratio" ("O/E Ratio") refers to the frequency of CpG

dinucleotides within a particular DNA sequence, and corresponds to the [number of CpG sites / (number of C bases X number of G bases)] X band length for each fragment.

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"CpG Island" refers to a contiguous region of genomic DNA that satisfies the criteria of (1) having a frequency of CpG dinucleotides corresponding to an "Observed/Expected Ratio" >0.6), and (2) having a "GC Content" >0.5. CpG islands are typically, but not always, between about 0.2 to about 1 kb in length. A CpG island sequence associated with a particular SEQ ID NO sequence of the present invention is that contiguous sequence of genomic DNA that encompasses at least one nucleotide of the particular SEQ ID NO sequence, and satisfies the criteria of having both a frequency of CpG dinucleotides corresponding to an Observed/Expected Ratio >0.6), and a GC Content >0.5.

"Methylation state" refers to the presence or absence of 5-methylcytosine ("5-mCyt") at one or a plurality of CpG dinucleotides within a DNA sequence.

"Hypermethylation" refers to the methylation state corresponding to an *increased* presence of 5-mCyt at one or a plurality of CpG dinucleotides within a DNA sequence of a test DNA sample, relative to the amount of 5-mCyt found at corresponding CpG dinucleotides within a normal control DNA sample.

"Hypomethylation" refers to the methylation state corresponding to a decreased presence of 5-mCyt at one or a plurality of CpG dinucleotides within a DNA sequence of a test DNA sample, relative to the amount of 5-mCyt found at corresponding CpG dinucleotides within a normal control DNA sample.

"Methylation assay" refers to any assay for determining the methylation state of a CpG dinucleotide within a sequence of DNA.

"MS.AP-PCR" (Methylation-Sensitive Arbitrarily-Primed Polymerase Chain Reaction) refers to the art-recognized technology that allows for a global scan of the genome using CG-rich primers to focus on the regions most likely to contain CpG dinucleotides, and described by Gonzalgo et al., Cancer Research 57:594-599, 1997.

"MethyLight" refers to the art-recognized fluorescence-based real-time PCR technique described by Eads et al., Cancer Res. 59:2302-2306, 1999.

"Ms-SNuPE" (Methylation-sensitive Single Nucleotide Primer Extension) refers to the art-recognized assay described by Gonzalgo & Jones, *Nucleic Acids Res.* 25:2529-2531, 1997.

"MSP" (Methylation-specific PCR) refers to the art-recognized methylation assay described by Herman et al. *Proc. Natl. Acad. Sci. USA* 93:9821-9826, 1996, and by US Patent No. 5,786,146.

"COBRA" (Combined Bisulfite Restriction Analysis) refers to the art-recognized methylation assay described by Xiong & Laird, *Nucleic Acids Res.* 25:2532-2534, 1997.

"MCA" (Methylated CpG Island Amplification) refers to the methylation assay described by Toyota et al., Cancer Res. 59:2307-12, 1999, and in WO 00/26401A1.

Overview

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The present invention provides for 103 DNA sequences (i.e., "marker sequences") having distinct methylation patterns in cancer, as compared to normal tissue. These methylation-altered DNA sequence embodiments correspond to 103 DNA fragments isolated from bladder and prostate cancer patients, and in many instances, represent novel sequences not found in the GenBank database. *None* of the instant sequence embodiments have previously been characterized with respect to their methylation pattern in human cancers including, but not limited to, those of the bladder and prostate. The significance of such methylation patterns lies in the value of altered fragments as potential prognostic, diagnostic and therapeutic markers in the treatment of human cancers.

Identification of Methylation-altered Marker Sequences in Genomic DNA

The MS.AP-PCR technique was used to scan the genomes of bladder or prostate cancer patients for DNA methylation changes relative to normal individuals, because the pattern is known to be highly conserved. A total of 103 DNA sequence embodiments (methylation-altered DNA sequences; "marker sequences") were isolated and characterized as having distinct methylation patterns in cancer, as compared to normal tissue.

Methods for the Identification of Marker Sequences in Genomic DNA. There are a variety of art-recognized genome scanning methods that have been used to identify altered methylation sites in cancer cells. For example, one method involves restriction landmark genomic scanning (Kawai et al., Mol. Cell. Biol. 14:7421-7427, 1994), another involves MCA (methylated CpG island amplification; Toyota et al., Cancer Res. 59:2307-12, 1999), and yet another involves MS.AP-PCR (Methylation-Sensitive Arbitrarily-Primed Polymerase Chain Reaction; Gonzalgo et al., Cancer Res. 57:594-599, 1997), which allows for a global scan of the genome using CG-rich primers to focus on the regions most likely to contain CpG dinucleotides. The MS.AP-PCR technique used in the present invention is a rapid and efficient method to screen ("scan") for altered methylation patterns in genomic DNA and to isolate specific sequences associated with these changes.

Briefly, genomic DNA from the tissue of bladder or prostate cancer patients was prepared using standard, art-recognized methods. Restriction enzymes (e.g., HpaII) with different sensitivities to cytosine methylation in their recognition sites were used to digest these genomic DNAs prior to arbitrarily primed PCR amplification with GC-rich primers. Fragments that showed differential methylation (e.g., hypermethylation or hypomethylation, based on the methylation sensitivity of the restriction enzyme, or upon DNA sequence analysis or Ms-SNuPE analysis; Gonzalgo & Jones, Nucleic Acids Res 25:2529-2531, 1997) were cloned and sequenced after resolving the PCR products on high-resolution polyacrylamide gels. The cloned fragments were used as probes for Southern blot analysis to

confirm differential methylation of these regions in the tissue. Methods for DNA cloning, sequencing, PCR, high-resolution polyacrylamide gel resolution and Southern blot analysis are well known by those of ordinary skill in the relevant art.

Results. A total of 500 DNA fragments that underwent either hypermethylation (an increase in the level of methylation relative to normal) or hypomethylation (a decrease in the level of methylation relative to normal) were isolated from the scanned patients genomic DNA. A total of 178 of these fragments were sequenced, of which 103 were novel in that they corresponded to DNA loci whose methylation pattern had not previously been characterized. The corresponding sequences are disclosed as [SEQ ID NOS:1-103], wherein for certain sequences, the letter "n" refers to an undetermined nucleotide base.

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Novel marker sequences identified by MS.AP-PCR. Table I shows an overall summary of methylation patterns and sequence data corresponding to the 103 DNA fragments identified by MS.AP-PCR. A total of 103 fragments were sequenced following identification as becoming either hypermethylated (gain of methylation; noted as having a hypermethylation pattern) or hypomethylated (loss of methylation; noted as having a hypomethylation pattern) relative to normal tissue. For the fragments of each category, the "Average GC Content" is shown, calculated as (number of C bases + number of G bases)/band length for each fragment, as well as the average Observed/Expected Ratio ("O/E Ratio"), calculated as [number of CpG sites/(number of C bases X number of G bases)] X band length for each fragment. Additionally, the percent of fragments that qualify as CpG islands is listed, and corresponds to the percentage of all fragments within each category that have sequence compositions that satisfy the criteria of having a "GC Content" >0.5 and an "O/E Ratio" >0.6.

Thus, of these 103 fragments identified by MS.AP-PCR, 60 showed hypermethylation (Table I, upper row; Table II, [SEQ ID NOS:1-60]) while 43 showed hypomethylation (Table I, lower row; Table II, [SEQ ID NOS:61-103]). Moreover, 55 (43 hypermethylated, and 12 hypomethylated) of the 103 fragments correspond to CpG islands (*i.e.*, fulfill the criteria of a GC content >0.5 and an Observed/Expected Ratio >0.6;), whereas the other 48 (17 hypermethylated and 31 hypomethylated) fragments do not meet the criteria for CpG islands (*see* Table II).

TABLE I. Summary of 103 DNA Fragments Identified by MS.AP-PCR

DNA Fragment Type	Methylation Pattern (relative to normal)	Number of Fragments (103 total)	Average GC Content	Average O/E Ratio	Percent that correspond to CpG Islands
Hypermethylated Fragments	Hyper- methylation	60	0.54	0.72	72%
Hypomethylated Fragments	Hypo-methylation	43	0.52	0.48	28%

Table II shows a summary of methylation pattern and sequence data for each individual sequence embodiment ([SEQ ID NOS:1-103]), corresponding to the 103 DNA fragments identified by MS.AP-PCR. Data for the 103 fragments was divided into either hypermethylated ([SEQ ID NOS:1-60]) or hypomethylated ([SEQ ID NOS:61-103]) categories. Table II also lists, for each sequence embodiment, the corresponding "Fragment Name," fragment "Size" (in base pairs; "bp"), "GC Content," Observed/Expected Ratio ("O/E Ratio"), "Description" (i.e., as a CpG island if criteria are met), "Inventor Initials" (IDCM = Isabel D.C. Markl, JC = Jonathan Cheng, GL = Gangning Liang, HF = Hualin Fu, YT = Yoshitaka Tomigahara), "Cancer Source," and "Chromosome Match" to the GenBank database. A dash ("-") indicates that no GenBank chromosome match existed, or that only a low-scoring partial match was found. Averages of the "GC Content" and "O/E Ratio," along with the percent of fragments that are CpG islands, are listed after the last member of both the hypermethylated and hypomethylated categories.

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Therefore, the present invention provides for 103 DNA fragments and corresponding marker sequence embodiments (i.e., methylation-altered DNA sequences) that are useful in cancer prognostic, diagnostic and therapeutic applications.

Additionally, at least 55 of these 103 sequences correspond to CpG islands (based on GC Content and O/E ration); namely [SEQ ID NOS:2, 4, 6, 7, 9-16, 19, 20, 22-33, 35-43, 48, 51-55, 59, 60, 64, 71, 76, 78-81, 84 and 87-90]. Thus, based on the fact that the methylation state of a portion of a given CpG island is generally representative of the island as a whole, the present invention further encompassed the novel use of the 55 CpG islands associated with [SEQ ID NOS:2, 4, 6, 7, 9-16, 19, 20, 22-33, 35-43, 48, 51-55, 59, 60, 64, 71, 76, 78-81, 84 and 87-90] in cancer prognostic, diagnostic and therapeutic applications, where a CpG island sequence associated with the sequence of a particular SEQ ID NO is that contiguous sequence of genomic DNA that encompasses at least one nucleotide of the particular SEQ ID NO sequence, and satisfies the criteria of having both a frequency of CpG dinucleotides corresponding to an Observed/Expected Ratio >0.6, and a GC Content >0.5.

TABLE II. Summary of MS.AP-PCR Fragments Sequenced

Methylation			GC	O/E	Description	Inventor	Cancer	Chromosome	[SEQ
Pattern	Name	(bp)	Content	Ratio		Initials	Source	Matches	ID NO
Hyper-									
methylation	,								
Category	11-1A	510	0.44	0.74		IDCM	Bladder	-	1
	14-3B	313	0.58	0.74	CpG Island	IDCM	Bladder	2	2
	18-2B	165	0.57	0.45		IDCM	Bladder	7	3
1	24-1B	601	0.51	0.72	CpG Island	IDCM	Bladder	Xp11	4
	26-1B	801	0.48	0.56		IDCM	Bladder	-	5
	26-2C	204	0.50	0.63	CpG Island	IDCM	Bladder	-	6
	30-3D	205	0.55	1.25	CpG Island	IDCM	Bladder	14	7
	32-3E	597	0.57	0.10	<u> </u>	IDCM	Bladder	20q12-13.1	8
	34-2B	500	0.62	0.66	CpG Island	IDCM	Bladder	20	9
	34-4B	343	0.70	0.81	CpG Island	IDCM	Bladder	-	10

	Fragment	Size	GC	O/E	Description	Inventor	Cancer	Chromosome	[SEQ
Pattern	Name	(bp)	Content	Ratio		Initials	Source	Matches	ID NO]
	34-5D	291	0.62	0.96	CpG Island	IDCM	Bladder	9	11
	34-6A	266	0.64	0.93	CpG Island	IDCM	Bladder	-	12
1	35-1C	553	0.64	0.63	CpG Island	IDCM	Bladder	ā.	13
	36-2D	156	0.60	0.58	CpG Island	IDCM	Bladder	10	14
1	38-1A	300	0.70	0.80	CpG Island	IDCM	Bladder	10	15
	38-2B	196	0.56	0.89	CpG Island	IDCM	Bladder	15	16
1	7-8E	299 363	0.59	0.39 0.49		IDCM IDCM	Bladder	17q21-22	17 18
	83-4B 84-1D	322	0.54 0.55	0.49	CpG Island	IDCM IDCM	Bladder Bladder	7	19
	101-3E	255	0.57	0.83	CpG Island	IDCM	Bladder	17	20
	M1-5A	406	0.45	0.96	Cpo Island	IDCM	Bladder	1	21
	U2-8E	210	0.56	0.61	CpG Island	IDCM	Bladder	2	22
	U12-1A	310	0.56	0.81	CpG Island	IDCM	Bladder	2	23
1	U7-4A	305	0.59	0.80	CpG Island	IDCM	Bladder		24
1	NU9-5A	379	0.67	0.83	CpG Island	JC	Bladder		25
	3-17-8-B	625	0.48	0.72	CpG Island	GL	Bladder	18	26
1	4-10-4-A	499	0.55	0.30	CpG Island	GL	Bladder	7	27
	1-1-1-A	561	0.58	0.98	CpG Island	GL	Bladder	20	28
	3-17-8-A	717	0.50	0.68	CpG Island	GL	Bladder	17	29
	G145-H	280	0.50	1.10	CpG Island	GL	Bladder	11	30
	1-1-1-D	270	0.50	0.60	CpG Island	GL	Bladder	2	31
	1-1-1-C	347	0.65	1.25	CpG Island	GL	Bladder	-	32
j	G178-A	342	0.55	0.85	CpG Island	GL	Bladder	2	33
	34-A'	370	0.62	0.44		HF	Prostate	-	34
	34-D	213	0.53	0.74	CpG Island	HF	Prostate	2	35
	35-D	173	0.56	0.66	CpG Island	HF	Prostate	3	36
1	36-A	369	0.67	0.70	CpG Island	HF	Prostate	-	37
	40-A	123	0.60	1.16	CpG Island	HF	Prostate	5 16-242	38
	91-1	450	0.64	0.86	CpG Island	YT	Bladder	5 or 16q24.3	39
İ	93-2	593	0.51	0.68	CpG Island	YT	Bladder	Xp11	40
,	93-3	457	0.52	0.94	CpG Island	YT	Bladder	Xp22.1-22.3	41
	94-8	211	0.66	0.96	CpG Island	YT	Bladder		42
1	95-5	141	0.63	0.79	CpG Island	YT	Bladder	14	43
1	97-5	559	0.56	0.40		YT	Bladder	-	44
	98-1	433	0.46	0.96		YT	Bladder	1	45
	100-1	487	0.59	0.58		YT YT	Bladder	14 3	46 47
ļ	100-2 100-6	403 155	0.60 0.57	0.47 0.99	CpG Island	YT	Bladder Bladder	20	48
	4-2	256	0.57	0.40	CpG Island	YT	Bladder	7	49
	5-8	224	0.47	0.96	}	YT	Bladder	5	50
	6-4	313	0.70	0.82	CpG Island	ŶŤ	Bladder	_	51
÷*	7-6	385	0.70	0.88	CpG Island	YT	Bladder		52
	13-3	307	0.59	0.89	CpG Island	YT	Bladder	10	53
	15-2	182	0.62	0.92	CpG Island	YT	Bladder	13	54
	23-2	523	0.54	0.87	CpG Island	YT	Bladder	Xp22.1-22.3	55
	39-2	795	0.46	0.64		YT	Bladder	13	56
	40-2	438	0.62	0.51	(YT	Bladder	10	57
· .	41-3	611	0.47	0.70)	YT	Bladder	18	58
	105-4	291	0.58	0.71	CpG Island	YT	Bladder	5	59
1	107-8	226	0.53	0.96	CpG Island	YT	Bladder	11	60
AVERAGE	,	ļ	0.54	0.72	72% islands				
Нуро-		l							
methylation					ł		51		
Category	14-2B	580	0.55	0.51	1 .	IDCM	Bladder	2	61
	16-1B	633	0.56	0.39		IDCM	Bladder	-	62
1	18-1B	703	0.45	0.35	1	IDCM	Bladder	17	. 63

Methylation	Fragment	Size	GC	O/E	Description	Inventor	Cancer	Chromosome	[SEQ
Pattern	Name	(bp)	Content			Initials	Source	Matches	ID NO
	19-1B	420	0.66	0.87	CpG Island	IDCM	Bladder	-	64
	20-1B	496	0.61	0.59		IDCM	Bladder	-	65
	21-2C	637	0.60	0.33		IDCM	Bladder	9q34	66
	29-1A	595	0.55	0.27		IDCM	Bladder	Xp11.23	67
	29-2B	580	0.47	0.77		IDCM	Bladder	-	68
	32-1A	589	0.59	0.48		IDCM	Bladder	-	69
(34-1B	450	0.42	0.46		IDCM	Bladder	-	70
'	34-3B	432	0.70	0.61	CpG Island	IDCM	Bladder	-	71
	32-2B	748	0.47	0.24	_	IDCM	Bladder	2	72
	32-4B	599	0.57	0.15		IDCM	Bladder	20q12-13.1	73
	32-5B	614	0.58	0.20		IDCM	Bladder	-	74
,	33-1A	552	0.54	0.32		IDCM	Bladder	10	75
,	5-1E	501	0.61	1.04	CpG Island	IDCM	Bladder	-	76
	6-1A	826	0.55	0.36		IDCM	Bladder	22q13.32- 13.33	77
	7-5D	433	0.59	0.85	CpG Island	IDCM	Bladder	5	78
	8-7C	424	0.58	0.83	CpG Island	IDCM	Bladder	5	79
	30-6D	285	0.63	0.72	CpG Island	IDCM	Bladder	1	80
	66-2E ·	401	0.54	0.82	CpG Island	IDCM	Bladder	16	81
	78-1C	268	0.54	0.41	_	IDCM	Bladder	-	82
	97-2E	989	0.53	0.16		IDCM	Bladder	-	83
	M1-8C	250	0.64	0.99	CpG Island	IDCM	Bladder	- 1	84
	M2-5A	402	0.50	0.45		IDCM	Bladder	5	85
	M1-4P	595	0.43	0.41		IDCM	Bladder	-	86
	M12-10A	304	0.53	0.76	CpG Island	IDCM	Bladder	7	87
	M12-12C	296	0.51	0.64	CpG Island	IDCM	Bladder	17	88
	M2-8M	220	0.67	0.62	CpG Island	IDCM	Bladder	6q27	89
	NU4-3A	273	0.63	1.02	CpG Island	JC	Bladder	-	90
	NU5-2A	361	0.44	0.73	·	JC	Bladder	6q14.3-15	91
	88-5	462	0.62	0.39		YT	Bladder	-	92
	90-1	591	0.66	0.45	1	YT	Bladder	19	93
	91-3	279	0.58	0.45		YT	Bladder	5 or 16q24.3	94
	91-4	351	0.55	0.30		YT	Bladder	18q23	95
	91-7	171	0.61	0.59		YT	Bladder	11	96
	89-3	743	0.55	0.43	}	YT	Bladder	-	97
	94-2	589	0.53	0.41		YT	Bladder	22q13.31- 13.32	98
	94-3	538	0.53	0.49		YT	Bladder	5 or 18	99
'	94-4	486	0.61	0.57		YT	Bladder		100
	94-5	450	0.60	0.45	1	YT	Bladder	1p36.2-36.3	101
	94-6	292	0.58	0.32		YT	Bladder	8 or 9	102
	96-4	395	0.63	0.54		YT	Bladder	9	103
AVERAGE		Ĺ	0.52	0.48	28% islands				

Diagnostic and Prognostic Assays for Cancer. The present invention provides for diagnostic and prognostic cancer assays based on determination of the methylation state of one or more of the disclosed 103 methylation-altered DNA sequence embodiments. Typically, such assays involve obtaining a tissue sample from a test tissue, performing a methylation assay on DNA derived from the tissue sample, and making a diagnosis or prognosis based thereon.

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The methylation assay is used to determine the methylation state of one or a plurality of CpG dinucleotide within a DNA sequence of the DNA sample. According to the present invention, possible methylation states include *hypermethylation* and *hypomethylation*, relative to a normal state (*i.e.*, non-cancerous control state). Hypermethylation and hypomethylation refer to the methylation states corresponding to an *increased* or *decreased*, respectively, presence 5-methylcytosine ("5-mCyt") at one or a plurality of CpG dinucleotides within a DNA sequence of the test sample, relative to the amount of 5-mCyt found at corresponding CpG dinucleotides within a normal control DNA sample.

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A diagnosis or prognosis is based, at least in part, upon the determined methylation state of the sample DNA sequence compared to control data obtained from normal, non-cancerous tissue.

Methylation Assay Procedures. Various methylation assay procedures are known in the art, and can be used in conjunction with the present invention. These assays allow for determination of the methylation state of one or a plurality of CpG dinucleotides (e.g., CpG islands) within a DNA sequence. Such assays involve, among other techniques, DNA sequencing of bisulfite-treated DNA, PCR (for sequence-specific amplification), Southern blot analysis, use of methylation-sensitive restriction enzymes, etc.

For example, genomic sequencing has been simplified for analysis of DNA methylation patterns and 5-methylcytosine distribution by using bisulfite treatment (Frommer et al., *Proc. Natl. Acad. Sci. USA* 89:1827-1831, 1992). Additionally, restriction enzyme digestion of PCR products amplified from bisulfite-converted DNA is used, *e.g.*, the method described by Sadri & Hornsby (*Nucl. Acids Res.* 24:5058-5059, 1996), or COBRA (Combined Bisulfite Restriction Analysis) (Xiong & Laird, *Nucleic Acids Res.* 25:2532-2534, 1997).

COBRA. COBRA analysis is a quantitative methylation assay useful for determining DNA methylation levels at specific gene loci in small amounts of genomic DNA (Xiong & Laird, Nucleic Acids Res. 25:2532-2534, 1997). Briefly, restriction enzyme digestion is used to reveal methylation-dependent sequence differences in PCR products of sodium bisulfite-treated DNA. Methylation-dependent sequence differences are first introduced into the genomic DNA by standard bisulfite treatment according to the procedure described by Frommer et al. (Proc. Natl. Acad. Sci. USA 89:1827-1831, 1992). PCR amplification of the bisulfite converted DNA is then performed using primers specific for the interested CpG islands, followed by restriction endonuclease digestion, gel electrophoresis, and detection using specific, labeled hybridization probes. Methylation levels in the original DNA sample are represented by the relative amounts of digested and undigested PCR product in a linearly quantitative fashion across a wide spectrum of DNA methylation levels. In addition, this technique can be reliably applied to DNA obtained from microdissected paraffin-embedded tissue samples. Typical reagents (e.g., as might be found in a typical COBRA-based kit) for

COBRA analysis may include, but are not limited to: PCR primers for specific gene (or methylation-altered DNA sequence or CpG island); restriction enzyme and appropriate buffer; gene-hybridization oligo; control hybridization oligo; kinase labeling kit for oligo probe; and radioactive nucleotides. Additionally, bisulfite conversion reagents may include: DNA denaturation buffer; sulfonation buffer; DNA recovery regents or kit (e.g., precipitation, ultrafiltration, affinity column); desulfonation buffer; and DNA recovery components.

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Preferably, assays such as "MethyLight" (a fluorescence-based real-time PCR technique) (Eads et al., *Cancer Res.* 59:2302-2306, 1999), Ms-SNuPE (Methylation-sensitive Single Nucleotide Primer Extension) reactions (Gonzalgo & Jones, *Nucleic Acids Res.* 25:2529-2531, 1997), methylation-specific PCR ("MSP"; Herman et al., *Proc. Natl. Acad. Sci. USA* 93:9821-9826, 1996; US Patent No. 5,786,146), and methylated CpG island amplification ("MCA";Toyota et al., *Cancer Res.* 59:2307-12, 1999) are used alone or in combination with other of these methods.

MethyLight. The MethyLight assay is a high-throughput quantitative methylation assay that utilizes fluorescence-based real-time PCR (TaqMan ®) technology that requires no further manipulations after the PCR step (Eads et al., Cancer Res. 59:2302-2306, 1999). Briefly, the MethyLight process begins with a mixed sample of genomic DNA that is converted, in a sodium bisulfite reaction, to a mixed pool of methylation-dependent sequence differences according to standard procedures (the bisulfite process converts unmethylated cytosine residues to uracil). Fluorescence-based PCR is then performed either in an "unbiased" (with primers that do not overlap known CpG methylation sites) PCR reaction, or in a "biased" (with PCR primers that overlap known CpG dinucleotides) reaction. Sequence discrimination can occur either at the level of the amplification process or at the level of the fluorescence detection process, or both.

The MethyLight may assay be used as a quantitative test for methylation patterns in 'the genomic DNA sample, wherein sequence discrimination occurs at the level of probe hybridization. In this quantitative version, the PCR reaction provides for unbiased amplification in the presence of a fluorescent probe that overlaps a particular putative methylation site. An unbiased control for the amount of input DNA is provided by a reaction in which neither the primers, nor the probe overlie any CpG dinucleotides. Alternatively, a qualitative test for genomic methylation is achieved by probing of the biased PCR pool with either control oligonucleotides that do not "cover" known methylation sites (a fluorescence-based version of the "MSP" technique), or with oligonucleotides covering potential methylation sites.

The MethyLight process can by used with a "TaqMan®" probe in the amplification process. For example, double-stranded genomic DNA is treated with sodium bisulfite and subjected to one of two sets of PCR reactions using TaqMan® probes; e.g., with either

biased primers and TaqMan® probe, or unbiased primers and TaqMan® probe. The TaqMan® probe is dual-labeled with fluorescent "reporter" and "quencher" molecules, and is designed to be specific for a relatively high GC content region so that it melts out at about 10 °C higher temperature in the PCR cycle than the forward or reverse primers. This allows the TaqMan® probe to remain fully hybridized during the PCR annealing/extension step. As the Taq polymerase enzymatically synthesizes a new strand during PCR, it will eventually reach the annealed TaqMan® probe. The Taq polymerase 5' to 3' endonuclease activity will then displace the TaqMan® probe by digesting it to release the fluorescent reporter molecule for quantitative detection of its now unquenched signal using a real-time fluorescent detection system.

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Typical reagents (e.g., as might be found in a typical MethyLight-based kit) for MethyLight analysis may include, but are not limited to: PCR primers for specific gene (or methylation-altered DNA sequence or CpG island); TaqMan® probes; optimized PCR buffers and deoxynucleotides; and Taq polymerase.

Ms-SNuPE. The Ms-SNuPE technique is a quantitative method for assessing methylation differences at specific CpG sites based on bisulfite treatment of DNA, followed by single-nucleotide primer extension (Gonzalgo & Jones, Nucleic Acids Res. 25:2529-2531, 1997). Briefly, genomic DNA is reacted with sodium bisulfite to convert unmethylated cytosine to uracil while leaving 5-methylcytosine unchanged. Amplification of the desired target sequence is then performed using PCR primers specific for bisulfite-converted DNA, and the resulting product is isolated and used as a template for methylation analysis at the CpG site(s) of interest. Small amounts of DNA can be analyzed (e.g., microdissected pathology sections), and it avoids utilization of restriction enzymes for determining the methylation status at CpG sites. Typical reagents (e.g., as might be found in a typical Ms-SNuPE-based kit) for Ms-SNuPE analysis may include, but are not limited to: PCR primers for specific gene (or methylation-altered DNA sequence or CpG island); optimized PCR buffers and deoxynucleotides; gel extraction kit; positive control primers; Ms-SNuPE primers for specific gene; reaction buffer (for the Ms-SNuPE reaction); and radioactive nucleotides. Additionally, bisulfite conversion reagents may include: DNA denaturation buffer; sulfonation buffer; DNA recovery regents or kit (e.g., precipitation, ultrafiltration, affinity column); desulfonation buffer; and DNA recovery components.

MSP. MSP (methylation-specific PCR) allows for assessing the methylation status of virtually any group of CpG sites within a CpG island, independent of the use of methylation-sensitive restriction enzymes (Herman et al. Proc. Natl. Acad. Sci. USA 93:9821-9826, 1996; US Patent No. 5,786,146). Briefly, DNA is modified by sodium bisulfite converting all unmethylated, but not methylated cytosines to uracil, and subsequently amplified with primers specific for methylated versus unmethylated DNA. MSP requires only small quantities of DNA, is sensitive to 0.1% methylated alleles of a given CpG island locus, and

can be performed on DNA extracted from paraffin-embedded samples. Typical reagents (e.g., as might be found in a typical MSP-based kit) for MSP analysis may include, but are not limited to: methylated and unmethylated PCR primers for specific gene (or methylationaltered DNA sequence or CpG island), optimized PCR buffers and deoxynucleotides, and specific probes.

MCA. The MCA technique is a method that can be used to screen for altered methylation patterns in genomic DNA, and to isolate specific sequences associated with these changes (Toyota et al., Cancer Res. 59:2307-12, 1999). Briefly, restriction enzymes with different sensitivities to cytosine methylation in their recognition sites are used to digest genomic DNAs from primary tumors, cell lines, and normal tissues prior to arbitrarily primed PCR amplification. Fragments that show differential methylation are cloned and sequenced after resolving the PCR products on high-resolution polyacrylamide gels. The cloned fragments are then used as probes for Southern analysis to confirm differential methylation of these regions. Typical reagents (e.g., as might be found in a typical MCA -based kit) for MCA analysis may include, but are not limited to: PCR primers for arbitrary priming Genomic DNA; PCR buffers and nucleotides, restriction enzymes and appropriate buffers; gene-hybridization oligos or probes; control hybridization oligos or probes.

Kits for Detection of Methylated CpG-containing Nucleic Acid. The reagents required to perform one or more art-recognized methylation assays (including those identified above) are combined with primers or probes comprising the sequences of SEQ ID NOS:1-103, or portions thereof, to determine the methylation state of CpG-containing nucleic acids. For example, the MethyLight, Ms-SNuPE, MCA, COBRA, and MSP methylation assays could be used alone or in combination, along with primers or probes comprising the sequences of SEQ ID NOS:1-103, or portions thereof, to determine the methylation state of a CpG dinucleotide within a genomic sequence corresponding to SEQ ID NOS:1-103, or to CpG island sequences associated with sequences of SEQ ID NOS:1-103, where the CpG island sequence associated with the sequence of the particular SEQ ID NO is that contiguous sequence of genomic DNA that encompasses at least one nucleotide of the particular SEQ ID NO sequence, and satisfies the criteria of having both a frequency of CpG dinucleotides corresponding to an Observed/Expected Ratio >0.6, and a GC Content >0.5.

We claim:

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1. A diagnostic or prognostic assay for cancer, comprising:

- (a) obtaining a tissue sample from a test tissue;
- (b) performing a methylation assay on DNA derived from the tissue sample, wherein the methylation assay determines the methylation state of a CpG dinucleotide within a DNA sequence of the DNA, and wherein the DNA sequence is a sequence selected from the group consisting of sequences of SEQ ID NOS:1-103, sequences having a nucleotide sequence at least 90% identical to sequences of SEQ ID NOS:1-103, CpG island sequences associated with sequences of SEQ ID NOS:1-103, CpG island sequences of SEQ ID NOS:1-103, and combinations thereof, wherein the CpG island sequence associated with the sequence of the particular SEQ ID NO is that contiguous sequence of genomic DNA that encompasses at least one nucleotide of the particular SEQ ID NO sequence, and satisfies the criteria of having both a frequency of CpG dinucleotides corresponding to an Observed/Expected Ratio >0.6, and a GC Content >0.5; and
 - (c) determining a diagnosis or prognosis based, at least in part, upon the methylation state of the CpG dinucleotide within the DNA sequence.
 - 2. The diagnostic or prognostic assay of claim 1 wherein the DNA sequence is a sequence selected from the group consisting of CpG island sequences associated with sequences of SEQ ID NOS:1-103, CpG island sequences associated with sequences having a nucleotide sequence at least 90% identical to sequences of SEQ ID NOS:1-103, and combinations thereof.
 - 3. The diagnostic or prognostic assay of claim 2 wherein the DNA sequence is a sequence selected from the group consisting of CpG island sequences associated with sequences of SEQ ID NOS: 2, 4, 6, 7, 9-16, 19, 20, 22-33, 35-43, 48, 51-55, 59, 60, 64, 71, 76, 78-81, 84 and 87-90, and combinations thereof.
 - 4. The diagnostic or prognostic assay of claim 1 wherein the methylation assay procedure is selected from the group consisting of MethyLight, MS-SNuPE, MSP MCA, COBRA, and combinations thereof.
 - 5. The diagnostic or prognostic assay of claim 1 wherein the methylation state of the CpG dinucleotide within the DNA sequence is that of hypermethylation, hypomethylation or normal methylation.
 - 6. The diagnostic or prognostic assay of claim 1 wherein the cancer is selected from the group consisting of bladder cancer, prostate cancer, colon cancer, lung cancer, renal cancer, leukemia, breast cancer, uterine cancer, astrocytoma, glioblastoma, and neuroblastoma.
 - 7. A kit useful for the detection of a methylated CpG-containing nucleic acid comprising a carrier means containing one or more containers comprising:

(a) a container containing a probe or primer which hybridizes to any region of a sequence selected from the group consisting of SEQ ID NOS:1-103, and sequences having a nucleotide sequence at least 90% identical to sequences of SEQ ID NOS:1-103; and

(b) additional standard methylation assay reagents required to affect detection of methylated CpG-containing nucleic acid based, at least in part, on the probe or primer.

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- 8. The kit of claim 7, wherein the additional standard methylation assay reagents are standard reagents for performing a methylation assay from the group consisting of MethyLight, MS-SNuPE, MSP MCA, COBRA, and combinations thereof.
- 9. The kit of claim 7, wherein the probe or primer comprises at least about 12 to 15 nucleotides of a sequence selected from the group consisting of SEQ ID NOS:1-103, and sequences having a nucleotide sequence at least 90% identical to sequences of SEQ ID NOS:1-103.
- 10. An isolated nucleic acid molecule comprising a methylated or unmethylated polynucleotide sequence selected from the group consisting of SEQ ID NO:1, SEQ ID NO:5, SEQ ID NO:6, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:13, SEQ ID NO:18, SEQ ID NO:24, SEQ ID NO:25, SEQ ID NO:32, SEQ ID NO:34, SEQ ID NO:37, SEQ ID NO:38, SEQ ID NO:42, SEQ ID NO:44, SEQ ID NO:51, SEQ ID NO:52, SEQ ID NO:62, SEQ ID NO:64, SEQ ID NO:65, SEQ ID NO:68, SEQ ID NO:69, SEQ ID NO:70, SEQ ID NO:71, SEQ ID NO:74, SEQ ID NO:76, SEQ ID NO:82, SEQ ID NO:83, SEQ ID NO:84, SEQ ID NO:86, SEQ ID NO:90, SEQ ID NO:92, SEQ ID NO:97, and SEQ ID NO:100.
 - 11. The nucleic acid of claim 10, wherein the nucleic acid is methylated.
 - 12. The nucleic acid of claim 10, wherein the nucleic acid is unmethylated.

PCT/US01/51652 WO 02/081749

SEQUENCE LISTING

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      position is 513 nucleotides
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                                                                    120
catagoaggt cottttgccc cggcagccat tottctgctc acaaggggct ggctctgggg
                                                                    180
acagggatgt ctttgtcatc agtgaccact aatccccctc ctcattggcc tccagggctg
                                                                    240
ctccccttca ctctcttggt tgaagttgta ggggctgagg ttaccctgag aaacacctgt
tcttggagcc catagaccca accttggaga tgcaggggga gccactggct gggctctgca
                                                                    360
ngtggggcca gctgatcccc anctgctggc acctccaggc atccacagag cttggagtcc
                                                                    420
cagccacatt tcctccttgg ccttagaggg agaggaagtc ctttgattgc ctagtccaag
                                                                    480
atccctttat ttcctgccct gggattatgg ggnagcaagc catgcccttc atgggaagct
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gttctccctt cctcggggtt gggtctggcc tcagctcggg caacagtcat gatgggc
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<400> 9

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cttctgct	ct tatttccaag	cttcgcgctt	tctacaaact	ccctgttgcc	ttgactttga	180
tttccago	ccg tggtgagggt	cagagtgaac	cccggcgcgc	tccccgacgg	catccccgca	240
caccagga	ata ggagaaattg	gagggcctgg	ggcctcgggc	tccgcagtcg	tcggaggaag	300
aacccacc	egc ggggtccgca	agggaaagtg	aagaggcccg	ggatttttcc	aaagcgctgg	360
ccaggaco	cc gaaggaaggg	gaggagtcac	ctgaagccgg	ggaaggcccc	ttgggtgctc	420
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gtttcccc	cca acttggcttc					500
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tgtccagg	gga gtgtccttca	ccaccacact	cctcacgtcc	aggcagtgat	cgacggcctg	180
geggeaco	ect cacageggge	ccatagcacg	gggccacaca	cgtcccctga	gcttagcctg	240
ggcacatt	eg tetgeegeeg	agggcttaag	ccagtctgca	gecegegeee	cgtcactcgg	300
acgcaagt	cc gtcgtccgct	ctgccacgcg	gccgctaagc	cga		343
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caaggacc	egc gegegegaag	gcgccgtagc	aagtgggcac	acaccagaca	ccaccccggc	120
gtgttccg	jcg ggagaagcca	gtgcacacat	cctcccgcaa	ggcggggttg	ccagtgcaac	180
acaggaat	cc tgccctttt	ctagaaaagc	cccctcccc	actttccctc	caatacactc	240
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	12 266					

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ggaggtt	cet gggaeteate tetgateeae egtettgegt tetetgggeg categaette	180								
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		266								
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tggtgtc	cacc atgegetete ecceggeace ggegaggega aaegtttege tagteeeegg	180								

gaggccc	ctc	ggtcagggca	gcagcatccc	tgcaccctct	ccgcaggtgg	tctccccgac	240		
gccacag	gtg	gccagcaggg	cgcgggtggg	ggcaggagcg	cctctcccct	gcccaggcct	300		
cccgctc	ctt	ctcggagcgc	tgtggcgggg	tggagagaca	gccttctaca	gctagtctag	360		
ctcggcg	cgg	ttcccgtctg	tggcctccta	atcccacage	cacagcgcct	tcctctaacc	420		
tccctcg	gtg	ggcttaaagc	ctcccgttcc	ttctgtctca	ttccttctgc	tecetecece	480		
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ggccgng	gcc	caa		•			553		
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ccaccgc	ccc	ctagacacgg	gtgaaaacct	gcctaaaagc	taactcaggc	agtgactcta	120		
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gcggggc	agg	cgggcgggcg	aagggagctc	cggntacgca	ganaacgcgg	agcgccccct	180		

toccaco	etge ge	gagggcat	cctgcccggg	ggaggaaagg	cgggagtccg	aggcgggtcg	240
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ctgacco	cgtg ct	tctctacc	ttcggaggtg	ggacagttgc	acgaagtgct	agttagaccg	120
gatcagt	ttgg aa	ıctgacgga	ggactgcaaa	gaagaaacta	aaatagacgt	cgaaagcctg	180
tcctcg	gcgt cg	rcaaa					196
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					gtggagctgg		60 120
•					gccccttcta		180
					agtgtatcag		240
	_				ctccagggga		299
cacatg	gece tg	geegggee	cccgggcagg	aycayycyay	aggtctgcgc	ggccgctaa	233
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					•	aacacagggc.	120
					gccacaagtc		180
					caggggcttg		240
- Lucalu	uguu et	-u-u-u-a-u					270

aaacaga	agga gggggggatt	ttaaatttca	gtccgtccaa	ccctgtagat	ctgctgtcct	300
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cga						363
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tttttca	aaag taaacgcttc	gggctgcagg	acactcagct	aagagcatca	ggggggcgcc	180
aagagg	caag gggcggggat	gggtggtggc	tcgcctcgtg	gcagaccgcc	cgcccgctcc	240
caagat	ccaa ctacgagctt	tttaactgca	gcaactttaa	tatacgctat	tggagctgga	300
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agacac	cgca gggagtcagg	teccegeega	caaatcggaa	gaggeetgeg	ggagttagcc	120
agataa	tget etecetgtee	tacccgtccc	caccaatttg	ccttttacct	gccgcagagc	180
ttgctt	gaac caaaggggtt	tgcggtcttc	tcctcctcaa	cttgcgatcc	ccaggccttc	240
gcgtcc	cgaa gccga					255
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<220>						
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<222> <223>	position is 6 min" refers to a		ined base			
<220>	•	•				
<221>		_				
<222> <223>	-		ined hase			

<220> <221> <222> <223>	unsure position is 18 nucleotides "n" refers to an undetermined base		
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tccctca	aaca tocaaaccaa cogagtgogt otgaggtgaa ategtgocag acttagagac	120	
ggctgc	cagg tttctctcaa gtcttggctt aacaaaagaa agcaaattac aaaaatggaa	180	
attttca	aaac tagegtteag tggtatteaa ategaegttt gggtagegea eaggeaeaga	240	
ccgcatt	tcgt gctattttgt gattaaaatg ataccaaaaa tacctccttg ctttggtttt	300	
cgtcttd	cgaa aacgacttot ttoottotto taatttoooc ottacttttg ggagoggoaa	360	
acccct	gacc actctagaat tgctaacatt tggaccggcg tcgcaa	406	
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<220> <221> <222> <223>	221> unsure 222> position is 13 nucleotides		
<220> <221> <222> <223>			
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gcctcct	tgga gacttngggg agagggatag ccggntaaag ctcctgtcct ttctataggc	120				
ataagegggt ggtcaccaeg gattggggat eeegaateee tggeteeaga tagaettaat 180						
gaagaagcac ctggatccgg gccgcgncaa 210						
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	unsure position is 9 nucleotides "n" refers to an undetermined base					
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cctagtt	tece gaggteetnn aetaggteta gateaetggg taaaagaagg ggageggean 1	20
cacgtat	tggg gtaggegete teactactea catetegaga eetttgeegg egtagggetg 1	80
tccgggg	ggga acgacccgcc ttttccggta tcggttgtca tggcggcgcc cagcccagcc	40
tggtttt	tttc cggtagccaa ttgaactaac aaccccgttc cctttaggac taatctgtca 3	00
cgtcggd	cgca · 3	10
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(225)	ii lololo to un unactolminos paro	
<220> <221>	WASHED.	
	unsure position is 266 nucleotides	
<223>	•	
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<221>		
<222> <223>	•	
<220>		
<221>	unsure	

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<400> 24
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                                                                                    60
tctggactta ccanagcaat tccagccggt gggcgtttgg cagtcactta aggaggtagg
                                                                                   120
                                                                                  180
gaaagcagcg agcttcaccg ggcgggctac gatgagtagc atgacgggca gcagcagcag
                                                                                   240
ccaqcaaaag ccctcqcaaa gtgtccagct gctgcactgc cgcggggact cccacagcac
                                                                                   300
catgactagt tegtgegact etgeancane aaacggette egaggaacae angategegg
                                                                                    304
gggca
<210> 25
<211> 379
<212> DNA
<213> Homo sapiens
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<220>
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<221> unsure
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<221> unsure
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	gtgg ctgggacggc caaggccgcg gcttcccgcg tggggatgcg ctntggcgca	120
	gtcc cggcggggcc aggcgtttgt gggcgggtga cggggatcta gggcttccgc	180
	·	240
	ttcc tcttgggctg tctttncggg tttggactcg cctgccaggc tgtgtgcagg	
	getg cetetggeeg geaggegtee gggetgeagg tgggeeggea ggeaggtgtt	300
agcggga	aagg gagcacaggt agcgaggtgg gatcggcgac ctggctaggg tgtcggcaga	360
atggaat	tgcg cggccgcta	379
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cctntca	agte atecaaaace tteaggette	cagggaggtt	ttgctataat	tttctctaag	120
catgact	tgtt tctgggggag gggaaagggg	tggttgtatt	tactgaaaat	tcaaatcgaa	180
ataataa	aatg gccaaatttg gacacttacg	gacccaaaca	gttttgctca	cgccagagaa	240
accgaga	agca cagggettge gtgaageeta	tctcggcaga	aggcaacatt	ctaataaagc	300
ccgtgg	gaaa acagattaca ttttcgccat	gaataagtça	tgcagtgaaa	aatattgcct	360
acagcc	tgtc gacttatatt attatcacgt	ttttcaactc	ggcgtgagga	gggagaggag	420
tgttca	tatt tgactaggaa ttgcaggatc	gatgcaaact	ccagggcagc	agccagactg	480
gcatate	gtgg ggctctccgg ttactttctc	tgtatgtcgc	gggtgagagg	aacagcgagg	540
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caatcaa	actc	caataactga	gctgaagttt	ttgtttaaaa	agaaaaaaat	ctgataagtg	120
atgattt	tac	ctacttgtgg	acactagatt	tcaattagga	aggtttttt	aaacggcttt	180
ttgtaac	ette	gctgcaggaa	gcaggtttgt	ttctttttct	tttcttttta	agagaaggtg	240
tatttca	actg	gtgcaatggc	ttggcacctc	cggggcctgg	gaggacctca	gacctcccca	300
gccctgc	ggtt	tctccgtctt	caagaccaac	taggaagggt	caagcgggga	gagggagtgg	360
agggtca	aggt	gagatctcag	agctgccccg	geeggeeeee	gtctctttct	acctcctctt	420
ccagaga	aacc	ageggeteae	accettetea	acgcaggaca	tgctcggcgg	ccaaagccga	480
attctgc	caga	tatccatca					499
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	pos	ition is 39	nucleotide an undeterm				
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<222>	
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<221>	
	position is 80 nucleotides
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<222>	
<223>	F
\2237	in leters to an undetermined base
4000	
<220>	
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	;
<220>	
	unsure
<221>	
	position is 544 nucleotides
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ggctnca	atec cegtegagen cagggeetee etgttetnet agacatneea aggageeaac 120
tcctcc	grag aagttocoog offotot tatttocaag offogogoff totacaaact 180

ccctgttgcc ttgactttga tttccagccg tggtgagggt cagagtgaac cccggcggcc 240
tccccgacgg catccccgca caccaggata ggagaaattg gagggcctgg gcctcggctc 300
ccgcagtcgt cggaggaaga acccaccgcg gggtccccaa gggaaagtga agaggcccgg 360
gattttcca aagcgctgcc aggaccccga aggaagggga ggagtcacct gaagccgggg 420
aagctccttg ggtgctctcc ttggatcctt atgttcactg actttcgcga ngccccctgg 480
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tgcngaaatc attacactng c 561

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<212> DNA
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<220> <221> unsure

<222> position is 651 nucleotides

<223> "n" refers to an undetermined base

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                                                                              60
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ttaaaacqcc ctacagaaaa tctcggcgaa gtccccggag aactctggtt tctaagatca
getgggegca ettteteegg gaegteeett etteteggte teagegeett eetgeeetea
qccqcqccnq tnttgttttg gtggcaaact gaaataagaa atggaaatat attggccttt
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<213> Homo sapiens
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<223> "n" refers to an undetermined base
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<222> position is 6 nucleotides
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 <222> position is 29 nucleotides <223> "n" refers to an undetermined base
 <220>
 <221> unsure
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 <222> position is 36 nucleotides
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<222> position is 244 nucleotides
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<221>	unsure	
	position is 265 nucleotides	
	"n" refers to an undetermined base	
~223 2	in releas to an undecermined base	
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gnggmig	mila micggegatg gathemigna gantenggtg atggatatet geagaatteg	00
gcttagc	oggo ogogaacaaa gagogaacca aaggatgott ogaattttta aaacggaato	120
tataas	cca aatgcaggac tggtgactta aggagctgcg aagtctgatt taccgggcct	180
cccycac	cca aatycaggac tygtgactta aggagotyog aagtotgatt taoogggoot	200
actctcc	gace tgcccccac ccccagctca gggggacctt tttatcntga acgccagagc	240
tacnnac	ccaa gtcgggtggc cacnnccaaa .	270
	3003330330 0000000	
-010-		
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<220>		
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	unsure	
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<223>		

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<220> <221> <222> <223>	£					
<220> <221> <222> <223>	unsure position is 325 nucleotides "n" refers to an undetermined base					
<220> <221> <222> <223>	position is 331 nucleotides					
<220> <221> <222> <223>						
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ctcaac	catt tegegetetg etetgteege tggtttgtee etgeeeggtt eeteteeeg	120				
ggcctg	teag ecteegette tetggaggtt cetgggacte atetetgate eacegtettg	180				
cgttct	ctgg gegeategae ttetetecat ettegggete acteetgaet eeetegetge	240				
cgcccc	gggg gtttccacgc gtgtctctaa ccgcggccgc taagccgaat tctgcagata	300				
tccatc	acng aantetgeag anatneateg negaannnea eegeaet	347				
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<220> <221> <222>						

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accgga	aaaa accaggctgg gctgggcgcc cgccatgaca accgataccg gaaaaggcgg	120
gtcgtt	cccc ccggacagec ctacgccggc aaaggteteg agatgtgagt agtgagageg	180
cctacc	ccat acngtcggcc ggctcccctt cttttaccca gtgatctaga cctagtctag	240
gacete	ggga actaggaeca geeteeetee ttettggaga tetgaecete aggatteann	300

nnctt	tgete acgageteca accenaenca tecaaannne aa	342
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gcaga	gecet getecetetg cageeceage teceagegee etgggettte caggeacetg	120
tccgg	gtagg ggattgaggg ccgtggccag gcccgcactt tcctgctagc cgcagctggc	180
cacat	gecea tetgaceete egagttetee tetaaaaatg gggetgacag eegetaeete	240
acaaa	gtoca caccgggete aacccgntge ettecteece aacaggaete tgecaccete	300
cctca	ggatg cctgagggcc ccganctgca cctggccagc cantttgtga atgaggcctg	360
ngggg	cgntt	370
<210><211><211><212><213><220><221><222><221><222>	213 DNA Homo sapiens unsure	٠
<223>	"n" refers to an undetermined base	
<220> <221>	· unsure	

	position is 10 nucleotides "n" refers to an undetermined base	
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	cnan taaagcgatg cttcgaattt ttaaaacgga atctctgcac ccaaatgcag	6
gactggt	gac ttaaggaget gegaagtetg atttacegge ctactetega cetgeecece	12
accccca	aget caggggacet tttgtctgaa cgccagaget actgaccagg tcggggggcc	18
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.01.05		
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.000:		
<220>		
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cgagcgtgaa gtcaagggct gcggtggatg ggggtaaaan gcctcctcnt cccac	etgcct 120
geneegtett ggggtaacce ctanceccca eeeggngttn enetttaatg etc	173
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caaggtccgt gacccgcgga ggtgatgggg gggataggag agccccaggg accgc	cagagg 120
tgacacaate geoegeeegt cetecetege tgggageega tteageetgt geega	ageete 180
tecettegeg tgeetetgeg cacageggtg geacegeagg acteegggte ecced	ccggct 240
ctccatcggg aagccggcaa atgcgcttcc tcagccagac cgcggcgggg tgggg	ggeggg 300
gggggcggaa gttgaaatac tgggacagaa acacctgccc gtcccaaggg acgga	aaaact 360
ggatgccaa	369
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<220> <221> <222> <223>	-	tion is 87	nucleotides an undetermi				
<220> <221> <222> <223>	_	tion is 108	3 nucleotide an undetermi				
<400>	38	cccacttttn	ctttccccna	aatcccaaca	necassecaa	cagatatosa	60
					-		120
	acag	ggcgagccgg	gggccangcg	gggccgcgca	aaatetentg	tygttattt	
gtg							123
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<220> <221> <222> <223>	~	tion is 32	nucleotides an undetermi				
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agcacct	tgcc	cgaagatcag	cccaggaccc	aaaggaaagc	aggctccaag	ctccccggaa	120
gccaag	gaaa	ataggaaaac	atatcctgcc	ccggggacac	cttctggaac	tatgaccaca	180
tgcacti	tgac	cttccggaac	aatcaccgca	tgcacctgac	ctcccggaac	tgtcaccacc	240
gcgcgca	acct	gacctcccgg	cactgtcacg	accgcgcgca	cctgacctcc	cggcactgtc	300
atcacco	gcgc	gcacctcacc	tcccggaact	gtcaccaccg	cgcgcacctg	acctcccggc	360
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cccggca	actg	tcacgaccgc	gcgcacctca				450
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	_				_		-

aaatcccacc	aaacgtgcag	ctggcactcg	gccgcctttg	tttccttccc	ctagaataaa	120
actccgctgc	tttcccacgt	tcctggagca	gcagccggaa	taaagcgccc	atggccttgc	180
cctttgagtc	tcggaggatg	tttgccactc	caacaatgga	cttttaaata	attcaggggt	240
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cccgcctctg	ctcccaacac	cctacgtttt	tctcttcctc	ctcatttacg	tatttacaat	180
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coggatcccg togggtttca actoctattc cgaaggtcct ttctccccta atcacaacac	120
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                                                                      240
                                                                      300
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atttcaagcc cagccagagc ctaccaggcc aacattcccc aggettgtgc ctctccgagc
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                                                                      420
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                                                                      720
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                                                                      840
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agccacaaaa atctttgntg ctggattcna aagtaggcat tggggctggg atctgggctc
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.2.20		Jupicino					
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	ccc ttgtggcacc agaagctaca a				180			
gactcgg	gtaa ttgctaagtg ccagctctga a	agtctgtgat	tccgtggaaa	tccctttcaa	240			
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- 3		
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	• • • • • • • • • • • • • • • • • • • •	
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